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1 Title: Body mass index as a modifiable risk factor for type 2 diabetes: Refining and understanding
2 causal estimates using Mendelian randomisation.

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13 Running title: Body mass index and type 2 diabetes; Mendelian randomisation methods

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22 **ABSTRACT**

23 This study focused on resolving the relationship between body mass index (BMI) and type 2
24 diabetes. The availability of multiple variants associated with BMI offers a new chance to resolve
25 the true causal effect of BMI on T2D, however the properties of these associations and their
26 validity as genetic instruments need to be considered alongside established and new methods for
27 undertaking Mendelian randomisation. We explore the potential for pleiotropic genetic variants to
28 generate bias, revise existing estimates and illustrate value in new analysis methods. A two-
29 sample Mendelian randomisation (MR) approach with 96 genetic variants was employed using
30 three different analysis methods, two of which (MR-Egger and the weighted median) have been
31 developed specifically to address problems of invalid instrumental variables. We estimate an odds
32 ratio for type 2 diabetes per unit increase in BMI (kg/m^2) of between 1.19 and 1.38, with the most
33 stable estimate using all instruments and a weighted median approach (1.26 95%CI (1.17, 1.34)).
34 *TCF7L2*(rs7903146) was identified as a complex effect or pleiotropic instrument and removal of
35 this variant resulted in convergence of causal effect estimates from different causal analysis
36 methods. This indicated the potential for pleiotropy to affect estimates and differences in
37 performance of alternative analytical methods. In a real type 2 diabetes focused example, this
38 study demonstrates the potential impact of invalid instruments on causal effect estimates and the
39 potential for new approaches to mitigate the bias caused.

40 Observational studies have shown body mass index (BMI) to be associated with risk of type 2
41 diabetes as well as with a range of diabetes-related metabolic traits (1; 2). However, it is well
42 known that confounding, reverse causation and biases can generate such associations and that
43 even with careful study design, incorrect inference is possible (3). One approach to circumventing
44 these problems is to use genetic association results within a Mendelian randomization (MR)
45 framework (3; 4). In MR analyses, genetic variants act as proxies for an exposure in a manner
46 independent of confounders. If in addition the variants only affect an outcome of interest through
47 the chosen exposure, then they are said to be valid instrumental variables (IVs). This enables
48 evaluation of the causal effect of the exposure on the outcome, escaping some of the limitations of
49 observational epidemiology; (5).

50

51 Following the success of genome-wide association studies (GWASs), the number of MR analyses
52 using large numbers of mostly uncharacterized variants associated with complex health outcomes
53 or intermediates is rapidly increasing (6; 7). In the case of BMI, there are now 97 genetic variants
54 reliably associated and there are examples where multiple variants have been used as a
55 composite IV to estimate the causal impact of BMI on health (8). Although using many IVs can
56 increase the power of MR analyses, it brings with it the concern that enlarged sets of genetic
57 variants are more likely to contain invalid IVs due to violations of the assumptions necessary for
58 valid causal inference using traditional methods (9). In particular, horizontal pleiotropy – where a
59 genetic variant affects the outcome via more than one biological pathway (10) – is a concern.
60 Importantly, the properties of these associations and their validity as genetic instruments need to
61 be considered alongside established and new methods for undertaking Mendelian randomisation.

62

63 In response to the general issue of using multiple genetic variants in MR, Bowden *et al.* (9)
64 propose both MR-Egger regression, an approach developed from the original Egger regression
65 technique for assessing small study bias in meta-analysis and a weighted weighted median
66 approach (11) as alternatives to the standard MR analysis. The MR-Egger and weighted weighted
67 median approaches both operate using distinct, but critically weaker, versions of the IV
68 assumptions, and therefore have the potential to deliver robust causal effect estimates. The MR-

69 Egger method also provides a formal statistical test as to whether or not the average pleiotropic
70 effect of the genetic variants is equal to zero (9).

71

72 **Research Design and Methods**

73 With increasing evidence for multiple biological pathways underlying type 2 diabetes (12; 13) and
74 increasing numbers of genetic variants available as IVs for BMI, we set out to test the potential for
75 bias in causal estimates from MR using these state-of-the-art approaches. We compared results
76 from MR-Egger regression (9) and weighted weighted median (11) approaches to a traditional
77 inverse-variance weighted (IVW) method (which makes the strong assumption that all variants are
78 valid IVs) (14) in an investigation of the causal relationship between BMI and type 2 diabetes.
79 These methods all undertake two-sample Mendelian randomisation whereby the GWAS results for
80 a disease outcome are unified with those of an exposure of interest and together used to estimate
81 the causal impact of that exposure on disease. We used published data in a two-sample analysis
82 strategy taking SNP-exposure and SNP-outcome associations from different sources (15; 16).

83

84 The effect sizes for BMI-associated SNPs with associated standard errors from a mixed-sex cohort
85 of European ancestry were taken from the Genetic Investigation of ANthropometric Traits (GIANT)
86 consortium (17) along with results for type 2 diabetes from the DIAbetes Genetics Replication And
87 Meta-analysis (DIAGRAM) Consortium. To avoid sample overlap, GIANT estimates were re-
88 calculated in the absence of DIAGRAM cohorts yielding a maximum sample size at any given
89 locus of 189,079. To aid interpretation of the effects of BMI on type 2 diabetes, effect sizes were
90 transformed to BMI units prior to analysis, assuming one standard deviation (SD) = $4.5\text{kg/m}^{2(17)}$.

91 For the corresponding SNP-outcome association, we took odds ratios (ORs) and confidence
92 intervals from a GWAS meta-analysis conducted by the DIAGRAM Consortium. This genome-wide
93 meta-analysis includes data from 12,171 type 2 diabetes cases and 56,862 controls of mainly
94 European descent imputed at up to 2.5 million autosomal SNPs (DIAGRAMv3) (18). All but one
95 (rs4787491, *INO80E*) of the BMI-associated SNPs ($p < 5 \times 10^{-8}$) from GIANT had results listed in the
96 DIAGRAMv3 dataset so 96 SNPs with results in both datasets were taken forward for analysis.

97

98 SNP-exposure and SNP-outcome associations were combined using the three different
 99 approaches outlined above. All analyses were conducted in R 3.2.0 (19). First, an inverse-variance
 100 weighted (IVW) method was implemented to provide a weighted average of the causal effect
 101 estimates (14). This method assumes that all genetic variants (i.e. 100%) satisfy the IV
 102 assumptions (including zero pleiotropy) and uses weights that assume the gene-exposure
 103 association estimates are measured without error (the No Measurement Error (NOME)
 104 assumption).
 105
 106 Second, we performed MR-Egger regression (9), which assumes NOME but allows each variant to
 107 exhibit pleiotropy. MR-Egger estimates remain consistent only if the magnitude of the gene
 108 exposure associations across all variants are independent of their pleiotropic effects (the InSIDE
 109 assumption) (9). As recommended by Bowden *et al* (9), the extent to which pleiotropy was
 110 balanced across the set of instruments as a whole was visually assessed by plotting the causal
 111 effect estimates against their precision, using a funnel plot and checking for asymmetry (Figure
 112 1A). The NOME assumption was assessed for MR-Egger via an adaptation of the I^2 statistic (I^2_{GX})
 113 (20) and adjusted for by combining MR-Egger with the method of Simulation Extrapolation
 114 (SIMEX) (21). Using SIMEX, new data sets are created by simulating gene-exposure association
 115 estimates under increasing violations of NOME and recording the amount of attenuation in the
 116 estimate that occurs. The set of attenuated estimates are then used to extrapolate back to the
 117 estimate that would have been obtained if NOME had been satisfied.
 118
 119 Finally, a weighted weighted median estimation method was applied (11). The weighted median
 120 provides a consistent estimate of causal effect if at least 50% of the information in the analysis
 121 comes from variants that are valid IVs. For a more detailed description of the three methods
 122 applied, see Online Appendix (Supplementary methods). A leave-one-out permutation analysis
 123 was conducted across all methods to assess the influence of potentially pleiotropic SNPs on the
 124 causal estimates (22). In the case of the linear models (IVW and MR-Egger) two additional
 125 analyses were conducted (23; 24). Firstly, the extent to which the causal estimate from each SNP

126 in the set could be considered an outlier was assessed using studentized residuals. Secondly,
127 Cook's distance (25) was used as a measure of the aggregate impact of each SNP on the model.

128

129 **Results**

130 All three approaches provide evidence of a positive causal relationship between BMI and type 2
131 diabetes. This is demonstrated in Figure 1B where the slope of the lines show the causal effect
132 estimates as predicted by the IVW, MR-Egger and m weighted median approaches. Estimates
133 correspond to an OR for type 2 diabetes per unit increase in BMI (kg/m^2) of 1.19, 1.26 and 1.38 for
134 the IVW, weighted median and MR-Egger analyses, respectively and are in line with a previous
135 MR estimate of 1.27 (95%CI 1.18, 1.36) (2) (Table 1). Assessment of the NOME assumption with
136 respect to the MR-Egger estimate gave $I_{GX}^2=0.83$, suggesting an approximate 15% attenuation of
137 the causal estimate towards zero. Bias adjustment via SIMEX gave a corrected MR-Egger causal
138 estimate of 1.46 (95%CI 1.16, 1.84) for type 2 diabetes per unit increase in BMI (kg/m^2).

139

140 Considering the individual SNP-based contributions to MR analysis, there is one clear outlier in the
141 distribution of effects shown in Figure 1 and that is *TCF7L2*(rs7903146). *TCF7L2*(rs7903146)
142 shows an association with BMI that is in the opposite direction to the overall trend (and weak
143 relative to its effect on type 2 diabetes), resulting in a large negative causal estimate from this SNP
144 alone. The presence of at least some unbalanced pleiotropy was detected within the set of
145 variants, as reflected by the intercept estimate of -0.019 ($p=0.10$) in the MR-Egger analysis.

146

147 To illustrate the impact of *TCF7L2*(rs7903146) on causal estimates, we performed a sensitivity
148 analysis in which each SNP in turn was removed from the set in a leave-one-out permutation
149 analysis. We saw a shift in the causal estimates from the IVW (an increase) and MR-Egger (a
150 decrease) as a result of the removal of *TCF7L2*(rs7903146) but no difference in the estimate from
151 the weighted median approach (Table 1; Figure 2). The results of the leave-one-out permutation
152 analysis showed that the impact of removing *TCF7L2*(rs7903146) from the variant set on the IVW
153 and MR-Egger estimates was greater than that of removing almost any other variant, with the
154 exception of *FTO*(rs1558902) (Figure 2A & B). When *FTO*(rs1558902) was removed, causal

155 estimates from both the IVW and MR-Egger analysis decreased (Table 1; Figure 2). In this
 156 instance we also observed movement in the causal effect estimate from the weighted median
 157 (Table 1; Figure 2C). The estimate of the intercept from MR-Egger moved closer to zero following
 158 both the removal of *TCF7L2*(rs7903146) and *FTO*(rs1558902) (Figure 2D). *TCF7L2*(rs7903146)
 159 was also identified as an outlier in both IVW and MR-Egger (studentized residuals, Bonferroni
 160 corrected $p < 1 \times 10^{-19}$) but *FTO*(rs1558902) was not (Online Appendix (Supplementary Results,
 161 Figures S1A/B)). Calculation of Cook's distance showed both variants to have a disproportionate
 162 level of influence on the model compared to other variants in the set (Online Appendix
 163 (Supplementary Results, Figures S2A/B)) .
 164
 165 These results suggest *TCF7L2*(rs7903146) may be pleiotropic with respect to the outcome, i.e.
 166 that it influences type 2 diabetes through an alternative pathway (other than BMI). Evidence from
 167 existing literature supports this assertion as the type 2 diabetes risk increasing allele at
 168 *TCF7L2*(rs7903146) has been associated with both increased fasting glucose (26) and decreased
 169 BMI (17). Under the assumption that *TCF7L2*(rs7903146) demonstrates horizontal pleiotropy with
 170 respect to type 2 diabetes, we would expect its inclusion in the variant set to bias the causal
 171 estimate predicted by the IVW approach, but not that predicted by MR-Egger or the weighted
 172 median. Removing *TCF7L2*(rs7903146) from the variant set causes a slight shift in the causal
 173 estimates from the IVW and MR-Egger approaches, bringing them more in line with one another
 174 and also with the weighted median estimate which remained stable in this instance. Also of note is
 175 the reduction in the 95% confidence interval of the MR-Egger estimate following removal of the
 176 *TCF7L2*(rs7903146). This increase in precision following removal of a likely invalid instrument from
 177 the set is another potentially favourable quality of this estimator. The relatively small changes
 178 observed across all methods as a result of removing *TCF7L2*(rs7903146) are in line with the
 179 relatively weak effect of the SNP as shown in Figure 1B.
 180
 181 In contrast, the effect of removing *FTO*(rs1558902) is more noticeable. Regardless of the method
 182 used, removing this variant results in a lower causal estimate (Table 1; Figure 2). The substantial
 183 influence of *FTO*(rs1558902) was predicable given the strength of its effect relative to the other

184 variants (Figure 1B), though properties of this effect are not in line with other variants used to
185 instrument BMI as reported elsewhere for physical activity (27), thyroid function (28) and
186 depression (29). The concomitant increase in standard error associated with the estimates here
187 point towards increased uncertainty moving the estimates towards the null in the absence of
188 *FTO*(rs1558902). The weighted median appears robust, even to the removal of *FTO*(rs1558902),
189 as demonstrated by the relatively tight distribution of estimates returned from the leave-one-out
190 permutation analysis (Figure 2C). This is as expected given the tolerance of weighted median
191 approaches to outliers.

192

193 **Discussion**

194 By applying new analytical techniques to an old question – the causal relationship between BMI
195 and type 2 diabetes – we have explored the potential for invalid instruments to bias causal
196 estimates in MR. In this case where BMI is the exposure, the opportunity to use a large instrument
197 list in causal analyses presents both opportunity, through variance explained, but also cost,
198 through complications generated by instrument properties or methods employed. Results here
199 suggest that both *TCF7L2* and *FTO* appear to have genetic variation which predicts BMI reliably,
200 but for which associations with type 2 diabetes do not fully align with that for other variants (given
201 BMI effects and assumed causality).

202

203 For *TCF7L2*, only recently suggested to be associated with BMI directly (17), this is not surprising
204 and reinforces the important point that the validity of a specific method's MR estimate depends on
205 whether the genetic variants collectively satisfy its assumptions. In this case, it is possible that the
206 negative association with BMI observed in GIANT is the product of a form of bias where the risk of
207 type 2 diabetes is leading to effective treatment, health benefit and BMI reduction. This is
208 supported by the apparently causal negative relationship between type 2 diabetes and BMI seen in
209 a reciprocal analysis where BMI is the outcome of interest (Online Appendix (Supplementary
210 Results, Figure S3)), though is likely to be more a comment on study design than biological effect.

211

212 In this example, the use of recently derived methods (9; 11) designed to overcome problems
213 caused by directional pleiotropy, yields estimates which are more stable in the presence or
214 absence of potentially invalid instruments and confirm the likely magnitude of the average effect of
215 BMI on type 2 diabetes (i.e. from the most likely and stable estimate, an elevation of odds of
216 disease of ~26% for each additional unit of BMI). The comparison of results from different methods
217 for any set of potential instruments is important when assessing the reliability of causal inferences
218 and important for downstream interpretation. In this case, whilst it is impossible to model precisely,
219 one can estimate the hypothetical impact of an average population level change in lifecourse BMI
220 on type 2 diabetes. Given a population size of 64.1 million in the UK in mid 2013(30) and a
221 modelled prevalence of type 2 diabetes (including non-diagnosed cases) of 7.4%(31; 32), the
222 estimated reduction in odds for a 1kg/m² reduction would potentially yield a reduction in the
223 number of cases from ~4.7-3.6 million (a shift in prevalence to 5.6%).

224

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227 KHW and RCR. JB and SB contributed to method and script development. LJC conducted the
228 analysis and wrote the manuscript. JB prepared the (Online Appendix (Supplementary methods)).
229 NJT is the guarantor of this work and, as such, had full access to all the data in the study and
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242

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245

246 There are **no** conflicts to declare.

References

1. Prospective Studies C: Body-mass index and cause-specific mortality in 900 000 adults: collaborative analyses of 57 prospective studies. *The Lancet* 373:1083-1096
2. Holmes Michael V, Lange Leslie A, Palmer T, Lanktree Matthew B, North Kari E, Almoguera B, Buxbaum S, Chandrupatla Hareesh R, Elbers Clara C, Guo Y, Hoogeveen Ron C, Li J, Li Yun R, Swerdlow Daniel I, Cushman M, Price Tom S, Curtis Sean P, Fornage M, Hakonarson H, Patel Sanjay R, Redline S, Siscovick David S, Tsai Michael Y, Wilson James G, van der Schouw Yvonne T, FitzGerald Garret A, Hingorani AD, Casas Juan P, de Bakker Paul IW, Rich Stephen S, Schadt Eric E, Asselbergs Folkert W, Reiner Alex P, Keating Brendan J: Causal Effects of Body Mass Index on Cardiometabolic Traits and Events: A Mendelian Randomization Analysis. *The American Journal of Human Genetics* 2014;94:198-208
3. Davey Smith G, Hemani G: Mendelian randomization: genetic anchors for causal inference in epidemiological studies. *Human Molecular Genetics* 2014;23:R89-R98
4. Davey Smith G, Ebrahim S: 'Mendelian randomization': can genetic epidemiology contribute to understanding environmental determinants of disease? *International Journal of Epidemiology* 2003;32:1-22
5. Lawlor DA, Harbord RM, Sterne JAC, Timpson N, Davey Smith G: Mendelian randomization: using genes as instruments for making causal inferences in epidemiology. *Stat Med* 2008;27:1133-1163
6. Sleiman PMA, Grant SFA: Mendelian Randomization in the Era of Genomewide Association Studies. *Clinical Chemistry* 2010;56:723-728
7. Boef AGC, Dekkers OM, le Cessie S: Mendelian randomization studies: a review of the approaches used and the quality of reporting. *International Journal of Epidemiology* 2015;
8. Nordestgaard BG, Palmer TM, Benn M, Zacho J, Tybjaerg-Hansen A, Davey Smith G, Timpson NJ: The Effect of Elevated Body Mass Index on Ischemic Heart Disease Risk: Causal Estimates from a Mendelian Randomisation Approach. *PLoS Med* 2012;9:e1001212
9. Bowden J, Davey Smith G, Burgess S: Mendelian randomization with invalid instruments: effect estimation and bias detection through Egger regression. *International Journal of Epidemiology* 2015;44:512-525
10. Didelez V, Sheehan N: Mendelian randomization as an instrumental variable approach to causal inference. *Statistical Methods in Medical Research* 2007;16:309-330
11. Bowden J, Davey Smith G, Haycock PC, Burgess S: Consistent estimation in Mendelian randomization with some invalid instruments using a weighted median estimator. *Genetic Epidemiology* 2016;40:304-314
12. Yaghootkar H, Frayling T: Recent progress in the use of genetics to understand links between type 2 diabetes and related metabolic traits. *Genome Biol* 2013;14:1-7
13. Dimas AS, Lagou V, Barker A, Knowles JW, Mägi R, Hivert M-F, Benazzo A, Rybin D, Jackson AU, Stringham HM, Song C, Fischer-Rosinsky A, Boesgaard TW, Grarup N, Abbasi FA, Assimes TL, Hao K, Yang X, Lecoeur C, Barroso I, Bonnycastle LL, Böttcher Y, Bumpstead S, Chines PS, Erdos MR, Graessler J, Kovacs P, Morken MA, Narisu N, Payne F, Stancakova A, Swift AJ, Tönjes A, Bornstein SR, Cauchi S, Froguel P, Meyre D, Schwarz PEH, Häring H-U, Smith U, Boehnke M, Bergman RN, Collins FS, Mohlke KL, Tuomilehto J, Quertemous T, Lind L, Hansen T, Pedersen O, Walker M, Pfeiffer AFH, Spranger J, Stumvoll M, Meigs JB, Wareham NJ, Kuusisto J, Laakso M, Langenberg C, Dupuis J, Watanabe RM, Florez JC, Ingelsson E, McCarthy MI, Prokopenko I,

Investigators obotM: Impact of Type 2 Diabetes Susceptibility Variants on Quantitative Glycemic Traits Reveals Mechanistic Heterogeneity. *Diabetes* 2014;63:2158-2171

14. Burgess S, Butterworth A, Thompson SG: Mendelian Randomization Analysis With Multiple Genetic Variants Using Summarized Data. *Genetic Epidemiology* 2013;37:658-665

15. Burgess S, Scott R, Timpson N, Davey Smith G, Thompson S: Using published data in Mendelian randomization: a blueprint for efficient identification of causal risk factors. *Eur J Epidemiol* 2015:1-10

16. Pierce BL, Burgess S: Efficient Design for Mendelian Randomization Studies: Subsample and 2-Sample Instrumental Variable Estimators. *American Journal of Epidemiology* 2013;178:1177-1184

17. Locke AE, Kahali B, Berndt SI, Justice AE, Pers TH, Day FR, Powell C, Vedantam S, Buchkovich ML, Yang J, Croteau-Chonka DC, Esko T, Fall T, Ferreira T, Gustafsson S, Kutalik Z, Luan Ja, Magi R, Randall JC, Winkler TW, Wood AR, Workalemahu T, Faul JD, Smith JA, Hua Zhao J, Zhao W, Chen J, Fehrmann R, Hedman AK, Karjalainen J, Schmidt EM, Absher D, Amin N, Anderson D, Beekman M, Bolton JL, Bragg-Gresham JL, Buyske S, Demirkan A, Deng G, Ehret GB, Feenstra B, Feitosa MF, Fischer K, Goel A, Gong J, Jackson AU, Kanoni S, Kleber ME, Kristiansson K, Lim U, Lotay V, Mangino M, Mateo Leach I, Medina-Gomez C, Medland SE, Nalls MA, Palmer CD, Pasko D, Pechlivanis S, Peters MJ, Prokopenko I, Shungin D, Stancakova A, Strawbridge RJ, Ju Sung Y, Tanaka T, Teumer A, Trompet S, van der Laan SW, van Setten J, Van Vliet-Ostaptchouk JV, Wang Z, Yengo L, Zhang W, Isaacs A, Albrecht E, Arnlöv J, Arscott GM, Attwood AP, Bandinelli S, Barrett A, Bas IN, Bellis C, Bennett AJ, Berne C, Blagieva R, Bluher M, Bohringer S, Bonnycastle LL, Bottcher Y, Boyd HA, Bruinenberg M, Caspersen IH, Ida Chen Y-D, Clarke R, Warwick Daw E, de Craen AJM, Delgado G, Dimitriou M, Doney ASF, Eklund N, Estrada K, Eury E, Folkersen L, Fraser RM, Garcia ME, Geller F, Giedraitis V, Gigante B, Go AS, Golay A, Goodall AH, Gordon SD, Gorski M, Grabe H-J, Grallert H, Grammer TB, Graszler J, Gronberg H, Groves CJ, Gusto G, Haessler J, Hall P, Haller T, Hallmans G, Hartman CA, Hassinen M, Hayward C, Heard-Costa NL, Helmer Q, Hengstenberg C, Holmen O, Hottenga J-J, James AL, Jeff JM, Johansson A, Jolley J, Juliusdottir T, Kinnunen L, Koenig W, Koskenvuo M, Kratzer W, Laitinen J, Lamina C, Leander K, Lee NR, Lichtner P, Lind L, Lindstrom J, Sin Lo K, Lobbens S, Lohrbeier R, Lu Y, Mach F, Magnusson PKE, Mahajan A, McArdle WL, McLachlan S, Menni C, Merger S, Mihailov E, Milani L, Moayyeri A, Monda KL, Morken MA, Mulas A, Muller G, Muller-Nurasyid M, Musk AW, Nagaraja R, Nothen MM, Nolte IM, Pilz S, Rayner NW, Renstrom F, Rettig R, Ried JS, Ripke S, Robertson NR, Rose LM, Sanna S, Scharnagl H, Scholtens S, Schumacher FR, Scott WR, Seufferlein T, Shi J, Vernon Smith A, Smolonska J, Stanton AV, Steinthorsdottir V, Stirrups K, Stringham HM, Sundstrom J, Swertz MA, Swift AJ, Syvanen A-C, Tan S-T, Tayo BO, Thorand B, Thorleifsson G, Tyrer JP, Uh H-W, Vandenput L, Verhulst FC, Vermeulen SH, Verweij N, Vonk JM, Waite LL, Warren HR, Waterworth D, Weedon MN, Wilkens LR, Willenborg C, Wilsgaard T, Wojczynski MK, Wong A, Wright AF, Zhang Q, The LifeLines Cohort S, Brennan EP, Choi M, Dastani Z, Drong AW, Eriksson P, Franco-Cereceda A, Gadin JR, Gharavi AG, Goddard ME, Handsaker RE, Huang J, Karpe F, Kathiresan S, Keildson S, Kiryluk K, Kubo M, Lee J-Y, Liang L, Lifton RP, Ma B, McCarroll SA, McKnight AJ, Min JL, Moffatt MF, Montgomery GW, Murabito JM, Nicholson G, Nyholt DR, Okada Y, Perry JRB, Dorajoo R, Reinmaa E, Salem RM, Sandholm N, Scott RA, Stolk L, Takahashi A, Tanaka T, van't Hooft FM, Vinkhuyzen AAE, Westra H-J, Zheng W, Zondervan KT, The AC, The A-BMIWG, The CDC, The CC, The G, The I, The MI, The Mu TC, The MC, The PC, The ReproGen C, The GC, The International Endogene C, Heath AC, Arveiler D, Bakker SJL, Beilby J, Bergman RN, Blangero J, Bovet P, Campbell H, Caulfield MJ, Cesana G, Chakravarti A, Chasman DI, Chines PS, Collins FS, Crawford DC, Adrienne Cupples L, Cusi D, Danesh J, de Faire U, den Ruijter HM, Dominiczak AF, Erbel R, Erdmann J, Eriksson JG, Farrall M, Felix SB, Ferrannini E, Ferrieres J, Ford I, Forouhi NG, Forrester T, Franco OH, Gansevoort RT, Gejman PV, Gieger C, Gottesman O, Gudnason V, Gyllenstein U, Hall AS, Harris TB, Hattersley AT, Hicks AA, Hindorf LA, Hingorani AD, Hofman A, Homuth G, Kees Hovingh G, Humphries SE, Hunt SC, Hypponen E, Illig T, Jacobs KB, Jarvelin M-R, Jockel K-H, Johansen B, Jousilahti P, Wouter Jukema J, Jula AM, Kaprio J, Kastelein JJP, Keinanen-Kiukkaanniemi SM,

360 Kiemeney LA, Knekt P, Kooner JS, Kooperberg C, Kovacs P, Kraja AT, Kumari M, Kuusisto J,
 361 Lakka TA, Langenberg C, Le Marchand L, Lehtimäki T, Lyssenko V, Mannisto S, Marette A, Matise
 362 TC, McKenzie CA, McKnight B, Moll FL, Morris AD, Morris AP, Murray JC, Nelis M, Ohlsson C,
 363 Oldehinkel AJ, Ong KK, Madden PAF, Pasterkamp G, Peden JF, Peters A, Postma DS,
 364 Pramstaller PP, Price JF, Qi L, Raitakari OT, Rankinen T, Rao DC, Rice TK, Ridker PM, Rioux JD,
 365 Ritchie MD, Rudan I, Salomaa V, Samani NJ, Saramies J, Sarzynski MA, Schunkert H, Schwarz
 366 PEH, Sever P, Shuldiner AR, Sinisalo J, Stolk RP, Strauch K, Tonjes A, Tregouet D-A, Tremblay
 367 A, Tremoli E, Virtamo J, Vohl M-C, Volker U, Waeber G, Willemssen G, Witteman JC, Carola
 368 Zillikens M, Adair LS, Amouyel P, Asselbergs FW, Assimes TL, Bochud M, Boehm BO, Boerwinkle
 369 E, Bornstein SR, Bottinger EP, Bouchard C, Cauchi S, Chambers JC, Chanock SJ, Cooper RS, de
 370 Bakker PIW, Dedoussis G, Ferrucci L, Franks PW, Froguel P, Groop LC, Haiman CA, Hamsten A,
 371 Hui J, Hunter DJ, Hveem K, Kaplan RC, Kivimäki M, Kuh D, Laakso M, Liu Y, Martin NG, Marz W,
 372 Melbye M, Metspalu A, Moebus S, Munroe PB, Njølstad I, Oostra BA, Palmer CNA, Pedersen NL,
 373 Perola M, Perusse L, Peters U, Power C, Quertermous T, Rauramaa R, Rivadeneira F, Saaristo
 374 TE, Saleheen D, Sattar N, Schadt EE, Schlessinger D, Eline Slagboom P, Snieder H, Spector TD,
 375 Thorsteinsdottir U, Stumvoll M, Tuomilehto J, Uitterlinden AG, Uusitupa M, van der Harst P,
 376 Walker M, Wallaschofski H, Wareham NJ, Watkins H, Weir DR, Wichmann HE, Wilson JF, Zanen
 377 P, Borecki IB, Deloukas P, Fox CS, Heid IM, O'Connell JR, Strachan DP, Stefansson K, van Duijn
 378 CM, Abecasis GR, Franke L, Frayling TM, McCarthy MI, Visscher PM, Scherag A, Willer CJ,
 379 Boehnke M, Mohlke KL, Lindgren CM, Beckmann JS, Barroso I, North KE, Ingelsson E, Hirschhorn
 380 JN, Loos RJJ, Speliotes EK: Genetic studies of body mass index yield new insights for obesity
 381 biology. *Nature* 2015;518:197-206
 382
 383 18. Morris AP, Voight BF, Teslovich TM, Ferreira T, Segrè AV, Steinthorsdottir V, Strawbridge RJ,
 384 Khan H, Grallert H, Mahajan A, Prokopenko I, Kang HM, Dina C, Esko T, Fraser RM, Kanoni S,
 385 Kumar A, Lagou V, Langenberg C, Luan JA, Lindgren CM, Müller-Nurasyid M, Pechlivanis S,
 386 Rayner NW, Scott LJ, Wiltshire S, Yengo L, Kinnunen L, Rossin EJ, Raychaudhuri S, Johnson AD,
 387 Dimas AS, Loos RJJ, Vedantam S, Chen H, Florez JC, Fox C, Liu C-T, Rybin D, Couper DJ, Kao
 388 WHL, Li M, Cornelis MC, Kraft P, Sun Q, van Dam RM, Stringham HM, Chines PS, Fischer K,
 389 Fontanillas P, Holmen OL, Hunt SE, Jackson AU, Kong A, Lawrence R, Meyer J, Perry JRB,
 390 Platou CGP, Potter S, Rehnberg E, Robertson N, Sivapalaratnam S, Stančáková A, Stirrups K,
 391 Thorleifsson G, Tikkanen E, Wood AR, Almgren P, Atalay M, Benediktsson R, Bonnycastle LL,
 392 Burt N, Carey J, Charpentier G, Crenshaw AT, Doney ASF, Dorkhan M, Edkins S, Emilsson V,
 393 Eury E, Forsen T, Gertow K, Gigante B, Grant GB, Groves CJ, Guiducci C, Herder C, Hreidarsson
 394 AB, Hui J, James A, Jonsson A, Rathmann W, Klopp N, Kravic J, Krjutškov K, Langford C,
 395 Leander K, Lindholm E, Lobbens S, Männistö S, Mirza G, Mühleisen TW, Musk B, Parkin M,
 396 Rallidis L, Saramies J, Sennblad B, Shah S, Sigurðsson G, Silveira A, Steinbach G, Thorand B,
 397 Trakalo J, Veglia F, Wennauer R, Winckler W, Zabaneh D, Campbell H, van Duijn C, Uitterlinden
 398 AG, Hofman A, Sijbrands E, Abecasis GR, Owen KR, Zeggini E, Trip MD, Forouhi NG, Syvänen A-
 399 C, Eriksson JG, Peltonen L, Nöthen MM, Balkau B, Palmer CNA, Lyssenko V, Tuomi T, Isomaa B,
 400 Hunter DJ, Qi L, Wellcome Trust Case Control C, Investigators M, Consortium G, Consortium A-
 401 TD, Consortium SD, Shuldiner AR, Roden M, Barroso I, Wilsgaard T, Beilby J, Hovingh K, Price
 402 JF, Wilson JF, Rauramaa R, Lakka TA, Lind L, Dedoussis G, Njølstad I, Pedersen NL, Khaw K-T,
 403 Wareham NJ, Keinanen-Kiukkaanniemi SM, Saaristo TE, Korpi-Hyövälti E, Saltevo J, Laakso M,
 404 Kuusisto J, Metspalu A, Collins FS, Mohlke KL, Bergman RN, Tuomilehto J, Boehm BO, Gieger C,
 405 Hveem K, Cauchi S, Froguel P, Baldassarre D, Tremoli E, Humphries SE, Saleheen D, Danesh J,
 406 Ingelsson E, Ripatti S, Salomaa V, Erbel R, Jöckel K-H, Moebus S, Peters A, Illig T, de Faire U,
 407 Hamsten A, Morris AD, Donnelly PJ, Frayling TM, Hattersley AT, Boerwinkle E, Melander O,
 408 Kathiresan S, Nilsson PM, Deloukas P, Thorsteinsdottir U, Groop LC, Stefansson K, Hu F, Pankow
 409 JS, Dupuis J, Meigs JB, Altshuler D, Boehnke M, McCarthy MI: Large-scale association analysis
 410 provides insights into the genetic architecture and pathophysiology of type 2 diabetes. *Nature*
 411 *genetics* 2012;44:981-990
 412
 413 19. Team RC: R: A language and environment for statistical computing., 3.2.0 ed. Vienna, Austria,
 414 R Foundation for Statistical Computing, 2014
 415

20. Higgins JPT, Thompson SG, Deeks JJ, Altman DG: Measuring inconsistency in meta-analyses. *BMJ : British Medical Journal* 2003;327:557-560
21. Bowden J: Assessing the suitability of summary data for Mendelian randomization analyses using MR-Egger regression: the role of the I^2 statistic. Technical Report (available on request), University of Bristol 2015
22. Stone M: Cross-Validatory Choice and Assessment of Statistical Predictions. *Journal of the Royal Statistical Society Series B (Methodological)* 1974;36:111-147
23. Venables WN, Ripley BD: *Modern Applied Statistics with S*. New York, Springer, 2002
24. Fox J, Weisberg S: *An R Companion to Applied Regression*. Los Angeles, Sage, 2011
25. Cook RD: Detection of Influential Observation in Linear Regression. *Technometrics* 1977;19:15-18
26. Manning AK, Hivert M-F, Scott RA, Grimsby JL, Bouatia-Naji N, Chen H, Rybin D, Liu C-T, Bielak LF, Prokopenko I, Amin N, Barnes D, Cadby G, Hottenga J-J, Ingelsson E, Jackson AU, Johnson T, Kanoni S, Ladenvall C, Lagou V, Lahti J, Lecoeur C, Liu Y, Martinez-Larrad MT, Montasser ME, Navarro P, Perry JRB, Rasmussen-Torvik LJ, Salo P, Sattar N, Shungin D, Strawbridge RJ, Tanaka T, van Duijn CM, An P, de Andrade M, Andrews JS, Aspelund T, Atalay M, Aulchenko Y, Balkau B, Bandinelli S, Beckmann JS, Beilby JP, Bellis C, Bergman RN, Blangero J, Boban M, Boehnke M, Boerwinkle E, Bonnycastle LL, Boomsma DI, Borecki IB, Bottcher Y, Bouchard C, Brunner E, Budimir D, Campbell H, Carlson O, Chines PS, Clarke R, Collins FS, Corbaton-Anchuelo A, Couper D, de Faire U, Dedoussis GV, Deloukas P, Dimitriou M, Egan JM, Eiriksdottir G, Erdos MR, Eriksson JG, Eury E, Ferrucci L, Ford I, Forouhi NG, Fox CS, Franzosi MG, Franks PW, Frayling TM, Froguel P, Galan P, de Geus E, Gigante B, Glazer NL, Goel A, Groop L, Gudnason V, Hallmans G, Hamsten A, Hansson O, Harris TB, Hayward C, Heath S, Hercberg S, Hicks AA, Hingorani A, Hofman A, Hui J, Hung J, Jarvelin M-R, Jhun MA, Johnson PCD, Jukema JW, Jula A, Kao WH, Kaprio J, Kardina SLR, Keinanen-Kiukkaanniemi S, Kivimaki M, Kolcic I, Kovacs P, Kumari M, Kuusisto J, Kyvik KO, Laakso M, Lakka T, Lannfelt L, Lathrop GM, Launer LJ, Leander K, Li G, Lind L, Lindstrom J, Lobbens S, Loos RJF, Luan J, Lyssenko V, Magi R, Magnusson PKE, Marmot M, Meneton P, Mohlke KL, Mooser V, Morken MA, Miljkovic I, Narisu N, O'Connell J, Ong KK, Oostra BA, Palmer LJ, Palotie A, Pankow JS, Peden JF, Pedersen NL, Pehlic M, Peltonen L, Penninx B, Pericic M, Perola M, Perusse L, Peyser PA, Polasek O, Pramstaller PP, Province MA, Raikonen K, Rauramaa R, Rehnberg E, Rice K, Rotter JI, Rudan I, Ruokonen A, Saaristo T, Sabater-Lleal M, Salomaa V, Savage DB, Saxena R, Schwarz P, Seedorf U, Sennblad B, Serrano-Rios M, Shuldiner AR, Sijbrands EJG, Siscovick DS, Smit JH, Small KS, Smith NL, Smith AV, Stancakova A, Stirrups K, Stumvoll M, Sun YV, Swift AJ, Tonjes A, Tuomilehto J, Trompet S, Uitterlinden AG, Uusitupa M, Vikstrom M, Vitart V, Vohl M-C, Voight BF, Vollenweider P, Waeber G, Waterworth DM, Watkins H, Wheeler E, Widen E, Wild SH, Willems SM, Willemsen G, Wilson JF, Witteman JCM, Wright AF, Yaghootkar H, Zelenika D, Zemunik T, Zgaga L, Wareham NJ, McCarthy MI, Barroso I, Watanabe RM, Florez JC, Dupuis J, Meigs JB, Langenberg C: A genome-wide approach accounting for body mass index identifies genetic variants influencing fasting glycemic traits and insulin resistance. *Nat Genet* 2012;44:659-669
27. Richmond RC, Davey Smith G, Ness AR, den Hoed M, McMahon G, Timpson NJ: Assessing Causality in the Association between Child Adiposity and Physical Activity Levels: A Mendelian Randomization Analysis. *PLoS Med* 2014;11:e1001618
28. Taylor PN, Richmond R, Davies N, Sayers A, Stevenson K, Woltersdorf W, Taylor A, Groom A, Northstone K, Ring S, Okosieme O, Rees A, Nitsch D, Williams GR, Smith GD, Gregory JW, Timpson NJ, Tobias JH, Dayan CM: Paradoxical Relationship Between Body Mass Index and Thyroid Hormone Levels: A Study Using Mendelian Randomization. *J Clin Endocrinol Metab* 2016;101:730-738

- 473 29. Walter S, Kubzansky LD, Koenen KC, Liang L, Tchetgen Tchetgen EJ, Cornelis MC, Chang S-
474 C, Rimm E, Kawachi I, Glymour MM: Revisiting mendelian randomization studies of the effect of
475 body mass index on depression. *American Journal of Medical Genetics Part B: Neuropsychiatric*
476 *Genetics* 2015;168:108-115
477
- 478 30. Office of National Statistics(UK): Annual Mid-year Population Estimates, 2013. *Statistical*
479 *bulletin*, 2014
480
- 481 31. The National Cardiovascular Intelligence Network: Cardiovascular disease key facts. In
482 *Diabetes: Public Health England*, 2013
483
- 484 32. Adult obesity and type 2 diabetes. Public Health England, 2014
485

486 **Tables**

487 Table 1 – Estimates from the application of inverse-variance weighted, MR-Egger and weighted
 488 median Mendelian randomisation methodologies. Estimates represent the estimated causal effect
 489 of body mass index on type 2 diabetes.

| Method | Estimate | 95% CI | p-value |
|--|----------|----------------|------------------------|
| Complete variant set (n=96 SNPs) | | | |
| IVW | 1.20 | 1.09, 1.30 | 8.00×10^{-05} |
| MR-Egger | 1.39 | 1.14, 1.68 | 1.53×10^{-03} |
| MR-Egger ^(<i>a</i>) | -0.019 | -0.041, 0.004 | 0.10 |
| Weighted median | 1.26 | 1.17, 1.34 | 5.26×10^{-9} |
| <i>TCF7L2</i> (rs7903146) removed from the variant set (n=95 SNPs) | | | |
| IVW | 1.22 | 1.16, 1.28 | 1.49×10^{-11} |
| MR-Egger | 1.34 | 1.17, 1.51 | 9.71×10^{-06} |
| MR-Egger ^(<i>a</i>) | -0.011 | -0.024, -0.024 | 0.13 |
| Weighted median | 1.26 | 1.19, 1.32 | 3.29×10^{-10} |
| <i>FTO</i> (rs1558902) removed from the variant set (n=95 SNPs) | | | |
| IVW | 1.16 | 1.06, 1.27 | 1.31×10^{-03} |
| MR-Egger | 1.30 | 1.01, 1.65 | 0.04 |
| MR-Egger ^(<i>a</i>) | -0.012 | -0.038, 0.014 | 0.34 |
| Weighted median | 1.21 | 1.13, 1.28 | 6.81×10^{-08} |

490 Intercept coefficients MR-Egger^(*a*) represent the average pleiotropic effect of a genetic variant on
 491 type 2 diabetes risk. “IVW” refers to inverse variance weighted estimates, SNP refers to single
 492 nucleotide polymorphism.

493 **Figures**

494 Figure 1 – Genetic associations with body mass index (BMI) and type 2 diabetes from 96 variants measured in GIANT (17) and DIAGRAM (18),
495 respectively. *TCF7L2*(rs7903146) and *FTO*(rs1558902) are marked with a 'X' and labelled.

496 A - funnel plot of minor allele frequency corrected genetic associations with BMI (interpreted as instrument strength) against causal estimates based
497 on each genetic variant individually, where the causal effect is expressed in logs odds ratio of type 2 diabetes for each unit increase in BMI. The
498 overall causal estimates (β coefficients) of BMI on type 2 diabetes estimated by inverse-variance weighted (solid black line), MR-Egger (dashed black
499 line) and weighted median (dotted black line) methods are also shown. Grey solid line represent $x=0$, that is a causal estimate of zero.

500 B - scatter plot of genetic associations with type 2 diabetes against associations with BMI, with causal estimates (β coefficients) of BMI on type 2
501 diabetes estimated by inverse-variance weighted (solid line), MR-Egger (dashed line) and weighted median (dotted line) methods.

502

503 Figure 2 – Distributions of regression estimates resulting from leave-one-out permutation analysis. Solid line = estimate from main analysis (n=96
504 variants); dashed line = estimate with *TCF7L2*(rs7903146) removed; dotted line = estimate with *FTO*(rs1558902) removed.

505 A - Causal estimates (β coefficients) of BMI on type 2 diabetes estimated by an inverse-variance weighted method

506 B - Causal estimates (β coefficients) of BMI on type 2 diabetes estimated by MR-Egger

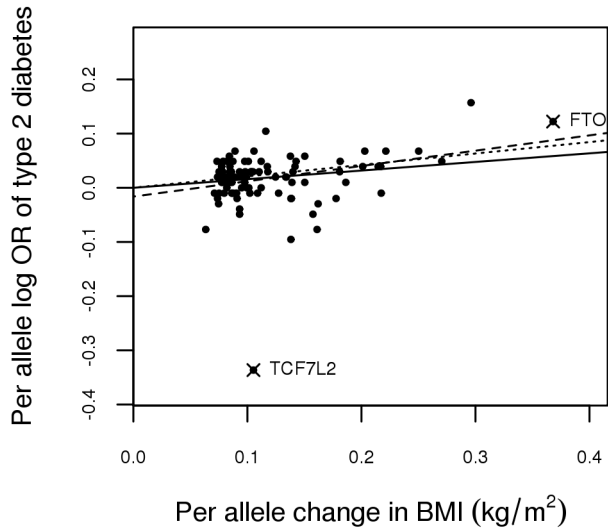
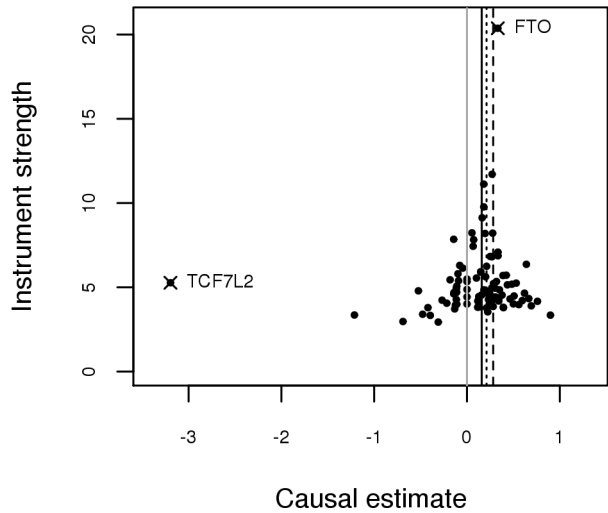
507 C - Causal estimates (β coefficients) of BMI on type 2 diabetes estimated by a weighted median method

508 D – Estimates of the intercept by MR-Egger

509

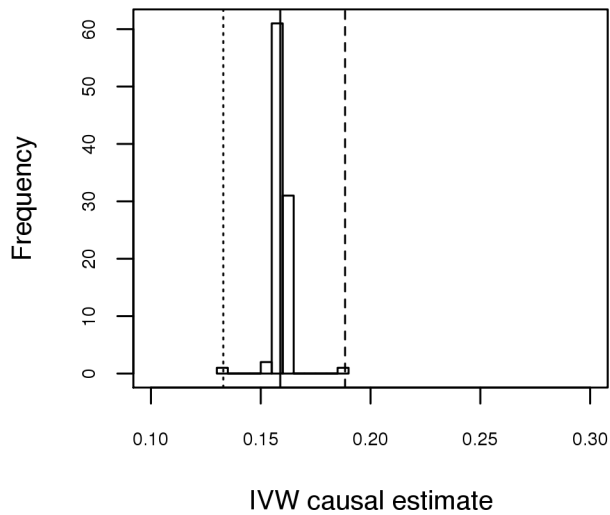
Figure 1

A

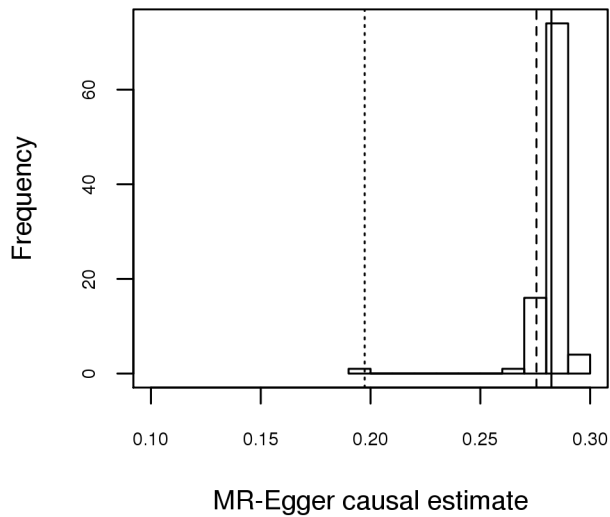


B

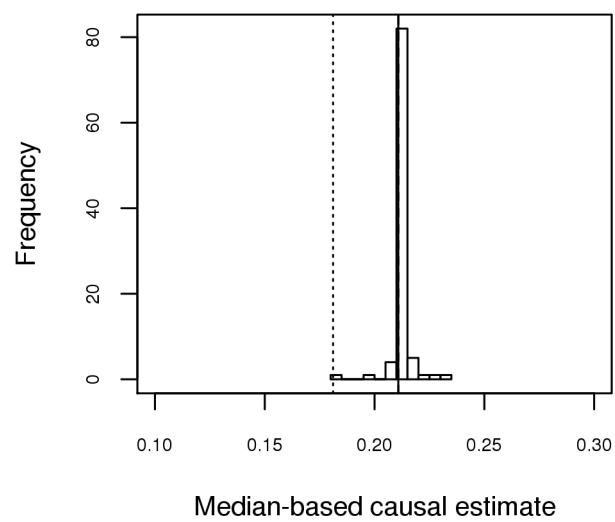
Figure 2



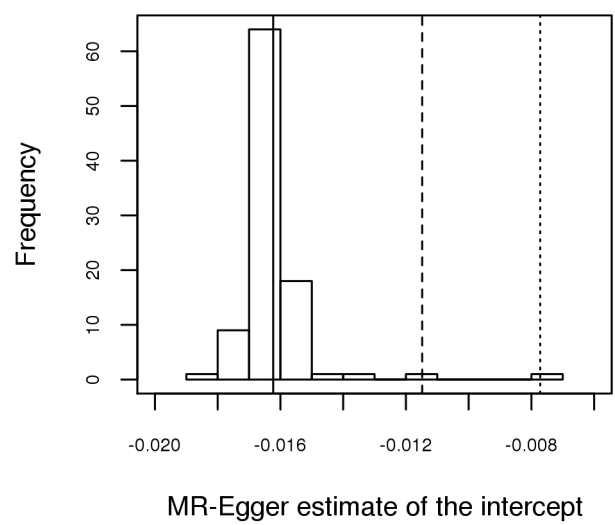
A



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C



D

Online Appendix

Supplementary methods

Mendelian randomization framework

Let $\hat{\Gamma}_j$ equal the gene-outcome association estimate for variant $j = 1, \dots, J$, with associated standard error σ_{Yj} . Let $\hat{\gamma}_j$ equal the gene-exposure association estimate for variant j , with associated standard error σ_{Xj} . Let the causal effect of the exposure on the outcome be denoted by β . An estimate for β based on variant j alone can be obtained via the ratio method as

$$\hat{\beta}_j = \frac{\hat{\Gamma}_j}{\hat{\gamma}_j}$$

Two forms for the variance of $\hat{\beta}_j$ are often used:

$$(i) \text{Var}(\hat{\beta}_j) = \frac{\sigma_{Yj}^2}{\hat{\gamma}_j^2}$$

$$(ii) \text{Var}(\hat{\beta}_j) = \frac{\sigma_{Yj}^2}{\hat{\gamma}_j^2} + \frac{\hat{\Gamma}_j^2 \sigma_{Xj}^2}{\hat{\gamma}_j^4},$$

Using either a first order (i) or second order (ii) Taylor series expansion. We use the variance from (i). This is equivalent to assuming that the gene-exposure association estimates are measured without error and is referred to as the No Measurement Error (NOME) assumption. NOME is equivalent to the assumption $\sigma_{Xj}^2 = 0$ for all j , so that $\hat{\gamma}_j = \gamma_j$ for all j .

The inverse variance weighted (IVW) method for the overall causal effect estimate

Let $w_j = 1 / \text{var}(\hat{\beta}_j)$ where $\text{var}(\hat{\beta}_j)$ is defined as in (i) under NOME. The inverse variance weighted (IVW) estimate for the causal effect is given by the standard meta-analytic formula

$$\frac{\sum_j w_j \hat{\beta}_j}{\sum_j w_j}.$$

The w_j terms derived under NOME are also referred to as ‘Toby Johnson’ weights. The IVW estimate assumes that all genetic variants satisfy the instrumental variable assumptions. If this is not true then it could give a biased estimate for β . The IVW estimate for β is consistent even if all genetic variants are invalid, provided that:

- Across all variants, the magnitude of the gene exposure associations are independent of their pleiotropic effects (the InSIDE assumption)
- NOME is satisfied
- The pleiotropic effects have zero mean

The weighted median method for the overall causal effect estimate

Let $\hat{\beta}_{(1)}, \dots, \hat{\beta}_{(J)}$ equal the J causal effect estimates ordered from smallest ($\hat{\beta}_{(1)}$) to largest ($\hat{\beta}_{(J)}$). Now define

$$w_{(j)}^* = \frac{w_j}{S_j}, \quad \text{where} \quad S_j = \sum_j w_j,$$

and equate $\hat{\beta}_{(j)}$ with a quantile, $p_{(j)}^w$, defined as

$$p_{(j)}^w = \frac{100}{S_j} \left(S_{(j)} - \frac{w_{(j)}}{2} \right).$$

$p_{(j)}^w$ represents the quantile from the weighted empirical distribution function of the ordered estimates $\hat{\beta}_{(1)}, \dots, \hat{\beta}_{(J)}$. The weighted median estimate, $\hat{\beta}_{WM}$ is defined as the 50th percentile of this weighted distribution. Typically the 50th percentile will lie between two estimates ($\hat{\beta}_{(l)}$ and $\hat{\beta}_{(m)}$, say), in which case $\hat{\beta}_{WM}$ is found by linear interpolation.

$\hat{\beta}_{WM}$ is a consistent estimate for β provided that at least 50% of the 'weight' making up S_j comes from genetic variants that are valid instruments.

The MR-Egger method for the overall causal effect estimate

The MR-Egger method performs a weighted linear regression of the gene-outcome coefficients on the gene-exposure coefficients:

$$\frac{\hat{\Gamma}_j}{\sigma_{Y_j}} = \frac{\beta_{0E}}{\sigma_{Y_j}} + \beta_{1E} \frac{\hat{\gamma}_j}{\sigma_{Y_j}}$$

The weights used are also derived under the NOME assumption. If all genetic variants are valid instruments, then $\beta_{0E} = 0$. The value of $\hat{\beta}_{0E}$ can be interpreted as an estimate of the average pleiotropic effect across the genetic variants. An intercept term that differs from zero is indicative of overall directional pleiotropy. The MR-Egger estimate for β , $\hat{\beta}_{1E}$, is consistent even if all genetic variants are invalid, provided that:

- Across all variants, the magnitude of the gene exposure associations are independent of their pleiotropic effects (the InSIDE assumption)

- NOME is satisfied.

If NOME is violated then the MR-Egger estimate of causal effect will be attenuated towards the null. We can assess the strength of NOME violation for MR-Egger through the I_{GX}^2

statistic: $I_{GX}^2 = \frac{Q-df}{Q}$, where $Q = \sum_{j=1}^J \frac{(\hat{\gamma}_j / \sigma_{Yj}^2 - \bar{\gamma})^2}{\sigma_{Xj}^2 / \sigma_{Yj}^2}$ and where $\bar{\gamma}$ equals the arithmetic mean of

the $\hat{\gamma}_j / \sigma_{Yj}^2$ terms. Specifically, the I_{GX}^2 statistic quantifies the proportion of the total variation

between the $\hat{\gamma}_j / \sigma_{Yj}^2$ terms that is due to 'true' variation between the γ_j / σ_{Yj}^2 terms.

Consequently, when NOME is satisfied $\hat{\gamma}_1, \dots, \hat{\gamma}_J = \gamma_1, \dots, \gamma_J$, I_{GX}^2 equals 1, and no attenuation occurs. When $I_{GX}^2 = 0.9$ we can expect the MR-Egger estimate to be only 90% of its value

had NOME been satisfied. A crude correction for NOME violation would be $\frac{\hat{\beta}_{1E}}{I_{GX}^2}$, however

this can be unstable as I_{GX}^2 can sometimes be estimated as zero, even when it is truly large.

We used the established method of Simulation Extrapolation (SIMEX) (1) instead, as implemented using the R package `simex()` (2). Under SIMEX, new data sets are created by simulating gene-exposure association estimates under increasing violations of NOME and recording the amount of attenuation in the estimate that occurs. The set of attenuated estimates are then used to extrapolate back to the estimate that would have been obtained if NOME had been satisfied.

Supplementary Results

Outlier analysis – Studentized residuals

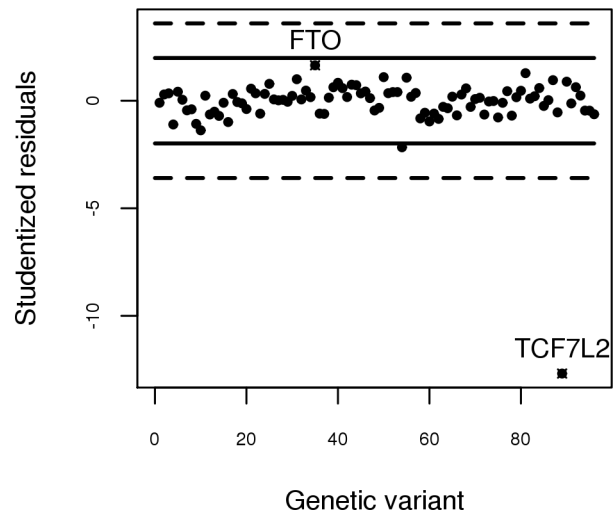


Figure S1A – Studentised residuals applied to the IVW method.

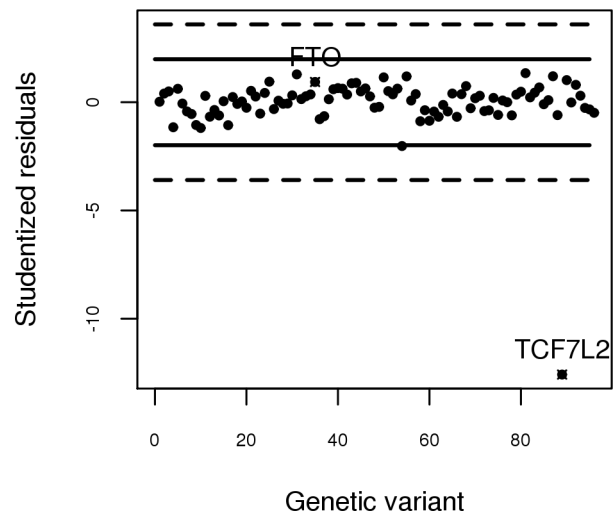


Figure S1B – Studentised residuals applied to the MR-Egger method.s

Outlier analysis – Cook's distance

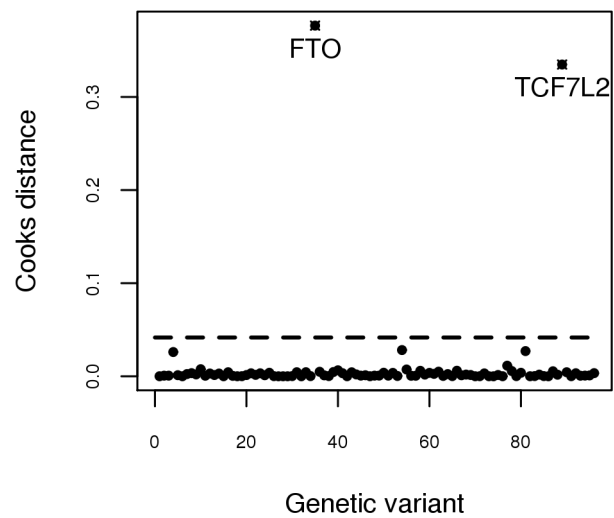


Figure S2A – Cook's distance applied to the IVW method.

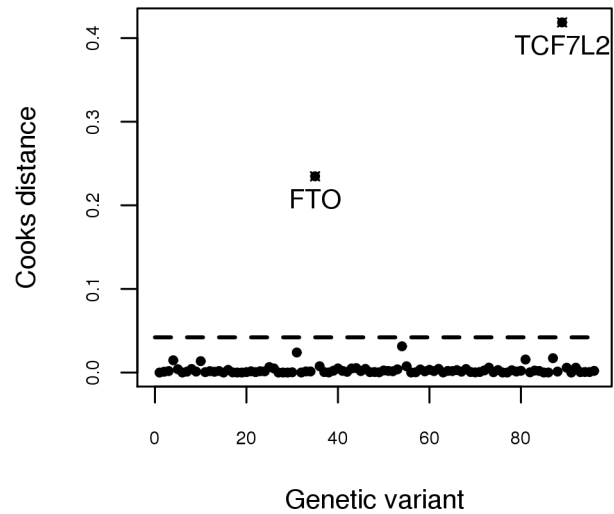


Figure S2B – Cook's distance applied to the MR-Egger method.

Reciprocal analysis of type 2 diabetes and BMI

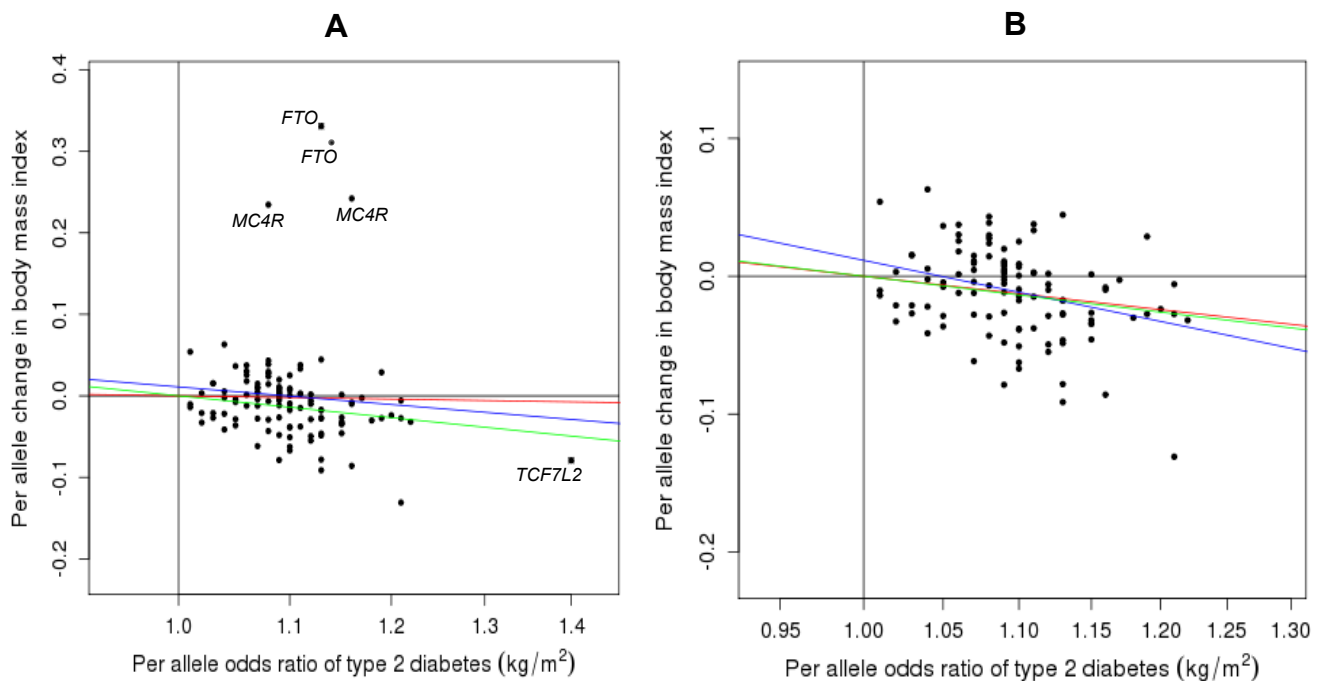


Figure S3 – MR-Egger analysis of the causal impact of type 2 diabetes on BMI.

A - scatter plot of genetic associations with BMI against associations with type 2 diabetes, with causal estimates (β coefficients) of type 2 diabetes on BMI estimated by inverse-variance weighted (red line), MR-Egger (blue line) and median-based (green line) methods. For this analysis, all 115 confirmed type 2 diabetes associated loci with OR not equal to 1 from Morris et al (2012)(3) downloaded from DIAGRAM <http://diagram-consortium.org/downloads.html> were used.

A - scatter plot of genetic associations with BMI against associations with type 2 diabetes, with causal estimates (β coefficients) of type 2 diabetes on BMI estimated by inverse-variance weighted (red line), MR-Egger (blue line) and median-based (green line) methods. For this analysis, 110 confirmed type 2 diabetes associated loci with OR not equal to 1 and no overlapping known BMI loci (excluding *FTO*, *MC4R* and *TCF7L2*) from Morris et al (2012)(3) were again used.

References

1. Cook JR, Stefanski LA: Simulation-Extrapolation Estimation in Parametric Measurement Error Models. *Journal of the American Statistical Association* 1994;89:1314-1328
2. Lederer W, Küchenhoff H: simex: SIMEX- and MCSIMEX-Algorithm for measurement error models., 2013
3. Morris AP, Voight BF, Teslovich TM, Ferreira T, Segrè AV, Steinthorsdottir V, Strawbridge RJ, Khan H, Grallert H, Mahajan A, Prokopenko I, Kang HM, Dina C, Esko T, Fraser RM, Kanoni S, Kumar A, Lagou V, Langenberg C, Luan Ja, Lindgren CM, Müller-Nurasyid M, Pechlivanis S, Rayner NW, Scott LJ, Wiltshire S, Yengo L, Kinnunen L, Rossin EJ, Raychaudhuri S, Johnson AD, Dimas AS, Loos RJJ, Vedantam S, Chen H, Florez JC, Fox C, Liu C-T, Rybin D, Couper DJ, Kao WHL, Li M, Cornelis MC, Kraft P, Sun Q, van Dam RM, Stringham HM, Chines PS, Fischer K, Fontanillas P, Holmen OL, Hunt SE, Jackson AU, Kong A, Lawrence R, Meyer J, Perry JRB, Platou CGP, Potter S, Rehnberg E, Robertson N, Sivapalaratnam S, Stančáková A, Stirrups K, Thorleifsson G, Tikkanen E, Wood AR, Almgren P, Atalay M, Benediktsson R, Bonnycastle LL, Burt N, Carey J, Charpentier G, Crenshaw AT, Doney ASF, Dorkhan M, Edkins S, Emilsson V, Eury E, Forsen T, Gertow K, Gigante B, Grant GB, Groves CJ, Guiducci C, Herder C, Hreidarsson AB, Hui J, James A, Jonsson A, Rathmann W, Klopp N, Kravic J, Krjutškov K, Langford C, Leander K, Lindholm E, Lobbens S, Männistö S, Mirza G, Mühleisen TW, Musk B, Parkin M, Rallidis L, Saramies J, Sennblad B, Shah S, Sigurðsson G, Silveira A, Steinbach G, Thorand B, Trakalo J, Veglia F, Wennauer R, Winckler W, Zabaneh D, Campbell H, van Duijn C, Uitterlinden AG, Hofman A, Sijbrands E, Abecasis GR, Owen KR, Zeggini E, Trip MD, Forouhi NG, Syvänen A-C, Eriksson JG, Peltonen L, Nöthen MM, Balkau B, Palmer CNA, Lyssenko V, Tuomi T, Isomaa B, Hunter DJ, Qi L, Wellcome Trust Case Control C, Investigators M, Consortium G, Consortium A-TD, Consortium SD, Shuldiner AR, Roden M, Barroso I, Wilsgaard T, Beilby J, Hovingh K, Price JF, Wilson JF, Rauramaa R, Lakka TA, Lind L, Dedoussis G, Njølstad I, Pedersen NL, Khaw K-T, Wareham NJ, Keinanen-Kiukkaanniemi SM, Saaristo TE, Korpi-Hyövälti E, Saltevo J, Laakso M, Kuusisto J, Metspalu A, Collins FS, Mohlke KL, Bergman RN, Tuomilehto J, Boehm BO, Gieger C, Hveem K, Cauchi S, Froguel P, Baldassarre D, Tremoli E, Humphries SE, Saleheen D, Danesh J, Ingelsson E, Ripatti S, Salomaa V, Erbel R, Jöckel K-H, Moebus S, Peters A, Illig T, de Faire U, Hamsten A, Morris AD, Donnelly PJ, Frayling TM, Hattersley AT, Boerwinkle E, Melander O, Kathiresan S, Nilsson PM, Deloukas P, Thorsteinsdottir U, Groop LC, Stefansson K, Hu F, Pankow JS, Dupuis J, Meigs JB, Altshuler D, Boehnke M, McCarthy MI: Large-scale association analysis provides insights into the genetic architecture and pathophysiology of type 2 diabetes. *Nature genetics* 2012;44:981-990